B1 could

proline, glutamic acid, serine and proline, are characteristically present in ustable proteins (Rogers et al., 1986, Science 234, 364-368).

Please replace the second full appearing on page 78, with the following paragraph:

B2

Insertional amino acid sequence variants of a protein of the invention are those in which one or more amino acid residues are introduced into a predetermined site in said protein. Insertions can comprise amino-terminal and/or carboxy-terminal fusions as well as intra-sequence insertions of single or multiple amino acids. Generally, insertions within the amino acid sequence will be smaller than amino or carboxyl terminal fusions, of the order of about 1 to 10 residues. Examples of amino- or carboxy-terminal fusion proteins or peptides include the binding domain or activation domain of a transcriptional activator as used in the yeast two-hybrid system, phage coat proteins, $(histidine)_6$-tag, glutathione S-transferase, protein A, maltose-binding protein, dihydrofolate reductase, Tag • 100 epitope (EETARFQPQPGPGYRS) (SEQ ID NO:42), c-myc epitope (EQKLISEEDL)(SEQ ID NO:43), FLAG® -epitope (DYKDDDK) (SEQ ID NO:44), lacZ, CMP (calmodulin-binding peptide), HA epitope (YPYDVPDYA) (SEQ ID NO:45), protein C epitope (EDQVDPRLIDGK) (SEQ ID NO:46) and VSV epitope (YTDIEMNRLGK) (SEQ ID NO:47).

Please replace the paragraph appearing on page 88 line 16 to page 89, line 22 with the following paragraph:

B3

The obtained *FL39* PCR fragment was purified, and cut with *NdeI* and *EcoRI* restriction enzymes. This fragment was cloned into the *NdeI* and *EcoRI* sites of pET derivative pRK172 (McLeod et al., 1987, EMBO J. 6, p729-736). The obtained *FL66* PCR fragment was purified, cut with *NcoI* and *BamHI* and cloned into the *NcoI* and *BamHI* sites of pET21d. FL66pET21d was transformed in *E. coli* BL21 (DE3). FL39pRK172 was co-transformed in *E. coli* BL21 (DE3) with pSBETa (Schenk et al., 1995 Biotechniques 19, p 196-200). PSBETa encoded the tRNA ^{UCU} that is low abundant tRNA in *E. coli*, corresponding to codons AGG and AGA (arginine). Because of the presence of an AGG AGA AGA sequence (SEQ ID NO:48) (Arg 5, Arg 6, Arg 7) at the beginning of *FL39* coding sequence, an increase of the tRNA ^{UCU} pool of *E. coli* is necessary for the translation of *FL39*. The FL66pET21d/BL21(DE3) and FL39pRK172, pSBETa/BL21(DE3) *E. coli* recombinant strains were grown in LB medium, supplemented respectively with 50 µg/ml ampicillin and 50 µg/ml ampicilline; 25 µg/ml kanamycine. The cells were grown at 37°C until the density

of the culture reached an A_{600nm}= 0.7. At this time point, 0.4 mM IPTG was added to induce the recombinant protein production. Cells were collected 3 hours later by centrifugation. The bacterial pellet from 250 ml culture was suspended in 10 ml lysis buffer (Tris.HCl pH7.5, 1 mM DTT, 1 mM EDTA, 1 mM PMSF and 0.1% Triton X-100) and submitted to three freeze/thaw cycles before sonication. Cell lysate was clarified by centrifugation 20 minutes at 8000 rpm. The pellet was collected, was suspended again in extraction buffer, the resulting suspension sonicated, and pellet collected by centrifugation 20 minutes at 8000 rpm. A third wash was performed the same way with Tris extraction buffer + 1M NaCl and a fourth wash with Tris extraction buffer. After the different washing steps, the pellet contains FL66 or FL39 protein at 90% homogeneity. The pellets were suspended in Laemli loading buffer (Laemmli, 1970, Nature 277, p 680-681) and FL66 and FL39 were further purified by SDS/ 12% polyacrylamide gel electrophoresis. The gel was stained in 0.025% coomassie brilliant blue R250 in water and destained in water. The strong band co-migrating at the 31 kDa molecular weight marker position was cut out of the gel with a scalpel. The polyacrylamide fragments containing FL66 or FL39 were lyopyhilized and reduced into powder. The rabbit immunization was performed in complete Freund adjuvant, sub-cutaneous, with these antigen preparations. One injection corresponded to 100 µg of protein. The boosting injections were performed with non-complete Freund adjuvant, sub-cutaneous. The obtained sera detected bands of the expected size in protein extracts prepared from 2-day-old actively dividing cell cultures. No signals were observed using the pre-immune sera.

Please replace the paragraph appearing at page 104, lines 3-20 with the following paragraph:

The full length ICK2-coding region was amplified by polymerase chain reaction (PCR) using the 5'-AGACCATGGCGGCGGTTAGGAG-3' (SEQ ID NO:41) and 5'-GGCGGATCCCGTCTTCTTCATGGATTC-3' (SEQ ID NO:10) primers and the pFL39 plasmid as template, introducing NcoI and BamHI restriction sites. The amplified fragment was cut with NcoI and BamHI and cloned between the NcoI and BamHI sites of PH35S (Hemerly et al., 1995), resulting into the 35SFL39 vector. The CaMV35S/ICK2/NOS cassette was released by EcoRI and XbaI and cloned blunt into the SmaI site of PGSV4 (Heourt et al, 1994). The resulting vector PGSFL39, was mobilized by the helper plasmid pRK2013 into *Agrobacterium tumefaciens* C58C1RifR harboring the plasmid pMP90. *A thaliana* plants ecotype Col-O were transformed by the floral dip method (Clough and Bent, 1998). Transgenic plants were obtained on kanamycin-containing media and later transferred

to soil for optimal seed production. For all analysis plants were grown *in vitro* with 16-h light/8-h dark illumination at 22 C on germination medium (GM, Valvekens *et al.*, 1988). Molecular analysis of the obtained transformants was performed by Northern as described by Jacqmard et al. (1999); and Western blotting and CDK kinase activity measurements as described by De Veylder et al. (1997).